Integrating GENomic prediction with GENe regulatory networks to optimize genetic value prediction : biological and statistical challenges

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IGEN: A 2 days seminar in December 2022

- **Biologists**
	- Christophe Perin
	- Nancy Terrier ٠
	- **Antoine Martin**
	- Gabriel Krouk ۰
	- ۰ \ddotsc
- Geneticists
	- **Vincent Segura**
	- Maud Fagny
	- **Renaud Rincent**
	- Stéphane Nicolas
	- **Celine Carilier Jacquin**
	- Mathilde Causse
	- **Emlie Millet**
	- Laurence Flori
	- Gabriel Krouk \bullet
	- \bullet ...
	- Statisticians
		- Andrea Rau \bullet
		- Sophie Lèbre
		- **Mickael Lucas** \bullet
		- Gabriel Krouk

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٠ ... Need to infer gene expression network/graph for addressing two biological objectives:

- **1** To gain insights into complex biological mechanisms involved in important processes, such as disease progress or growth
- ² To improve prediction of important phenotypes in genetic improvement context

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Towards a better understanding (genomic context)

To infer links/connections between genes for identifying biological mechanisms (such as key genes, functional modules, relations between network and a phenotype of interest, etc.)

Example:

How potassium and sodium fertilization impact biological mechanisms involved in response to water deficiency in Eucalyptus grandis ?

significance of the genes within the cluster purple (middle), and the associated enrichment Figure 1: Gene co-expression network (on left), bar plot representing the average gene map (on right) [\(Favreau et al., 2019\)](#page-22-1).

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Network inference

How to build co-expression network from gene expression data?

Data:

$$
X = \begin{pmatrix} X_{1,1} & X_{1,2} & \cdots & X_{1,n} \\ X_{2,1} & X_{2,2} & \cdots & X_{2,n} \\ \vdots & \vdots & \ddots & \vdots \\ X_{m,1} & X_{m,2} & \cdots & X_{m,n} \end{pmatrix}
$$

with $X_{i,j}$ the expression level of gene *j* for sample *i*

We want to infer **network/graph** where:

- Vertices: genes
- Edges: links between genes (gene-gene interactions)

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Network inference

What do we mean by links (gene-gene interactions)?

- Does it depend on biological question and/or experimental design?
- Does co-expression network aim at focusing on direct co-expression between genes? [\(Villa-Vialaneix et al., 2013;](#page-22-2) [Grimes et al., 2019\)](#page-22-3)

In the litterature

- Pearson-based correlation networks (relevance networks): marginal relationships between genes. Each pair of genes is considered alone: very dense networks, edges represent marginal connections not direct or causal
- Partial correlation based networks: direct relationships between genes. Correlation between two genes corrected for all other genes under investigation

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Network inference

Gene2 and Gene3 correlated but not dependent on each other

Gaussian Graphical Models (GGMs) [\(Lauritzen, 1996\)](#page-22-4) commonly used to estimate partial correlations

- Improve measurement of direct relations between gene expressions by accounting for the effect of all expression data
- \bullet More efficient for grouping together genes with a common function / more consistent to prior biological knowledge [\(Werhli et al., 2006;](#page-22-5) [Krumsiek et al., 2011;](#page-22-6) [Villa-Vialaneix et al., 2013\)](#page-22-2)

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Network inference: Gaussian Graphical Model

Let X_i be the *m*-vector (gene expression) of observed data for subject *i* such that

$$
X_i \sim \mathcal{N}_m(\mu, \Sigma), i = 1, \ldots n,
$$

with $\mu \in \mathbb{R}^m$ is the mean vector, Σ is the covariance matrix which is a positive semi-definite symmetric matrix, and $\Omega=\Sigma^{-1}\in\mathbb{R}^m\times\mathbb{R}^m$ is the precision matrix.

 \rightarrow Conditional independence implied by the form/structure of the precision matrix:

Gene j and Gene k are linked $\Leftrightarrow \Omega_{ik} > 0$.

Problem: When $n < m$, Σ is not full rank \Rightarrow can not be inverted

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Network inference: Gaussian Graphical Model

Various estimation techniques (from the review done by [Altenbuchinger et al. \(2020\)](#page-22-7)):

- More or less adapted for dealing with high-dimensional data: low to high differences observed
- More or less user friendly
- \Rightarrow \Rightarrow \Rightarrow Need guidelines for choosing the most adapted/t[o co](#page-9-0)[m](#page-11-0)[p](#page-8-0)a[re](#page-10-0)[th](#page-3-0)[e](#page-4-0)m

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Network evaluation

As most of co-expression networks in plants are Pearson-based correlation networks \Rightarrow Need to compare Pearson-based correlation network and partial correlation based network [\(Werhli et al., 2006;](#page-22-5) [Krumsiek et al., 2011\)](#page-22-6)

How to compare the inferred networks? How to evaluate their biological relevance ?

- Functional enrichment analysis for testing the biological relevance, detection of key genes, relevance of networks to the phenotype of interest, etc. [\(Villa-Vialaneix et al., 2013;](#page-22-2) [Lee et al., 2020\)](#page-22-8)
- To compare to a "reference" network (obtained from data base such as STRING protein-protein interactions database)
- \leftrightarrow Which statistical measures?
	- \bullet Co-expression Differential Network Analysis: to extract the common structure

[\(Grimes et al., 2019;](#page-22-3) [Peterson et al., 2020\)](#page-22-9)

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Is \ll functional understanding \gg relevant for prediction objectives, if it is the case how we take it into consideration ?

Idea: To use gene expression data or prior knowledge information into GS models

Genomic Selection (GS) model

$$
Y = \mu + \underbrace{X\beta}_{GEBV} + \varepsilon
$$

with $Y \in n \times 1$ the phenotype of interest, $X \in n \times p$ the marker matrix, $\beta \in p \times 1$ the marker effects, and $\varepsilon \sim \mathcal{N}_n(0, \sigma^2$ ldn). GEBV: Genomic Estimated Breeding Value

 \rightarrow Various statistical approaches for estimating marker effects $\hat{\beta}$ (Ridge regression, Bayesian Lasso (BayesB), BayesC, etc)

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Idea: To use gene expression data or prior knowledge information into GS models

$$
Y = \mu + X \underbrace{\beta}_{\text{prior knowledge information}} + \varepsilon
$$

Which type of information?

- From previous experimental studies (Co-expression networks, GO terms, GWAS results, selection signature,…) but may be not adequate with data at hand
- **•** From "physical" knowledge: markers belonging to the same gene

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Idea: To use gene expression data or prior knowledge information into GS models

$$
Y = \mu + X \underbrace{\beta}_{\text{prior knowledge information}} + \varepsilon
$$

"Although there are many databases that provide information on biochemical relationships under normal conditions, the available reference networks may be incomplete or inappropriate for the experimental condition or set of subjects under study" [\(Peterson et al., 2016\)](#page-22-10)

- \rightarrow Need to use statistical approaches integrating different degrees of fidelity/belief to the prior knowledge (to guard against mis-specification) [\(Stingo](#page-22-11) [et al., 2010;](#page-22-11) [Kundu et al., 2018;](#page-22-12) [Denis et al., 2022\)](#page-22-13)
- \leftrightarrow Need to use statistical approaches providing a trade-off between prior knowledge and computational complexity

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Idea: To use gene expression data or prior knowledge information into GS models

$$
Y = \mu + X \underbrace{\beta}_{\text{prior knowledge information}} + \varepsilon
$$

How to integrate those information into GS models?

Bayesian framework is a natural framework where prior knowledge may be specified via prior on regression coefficients (Bayesian fused and group Lasso (K_{Vung}) [et al., 2010\)](#page-22-14), Ising prior [\(Li and Zhang, 2010\)](#page-22-15)) Example: $\beta \sim \mathcal{N}_p(0, \Sigma)$ with Σ related to structure between variables specified via for instance by undirected graph (Graph Laplacian prior[\(Liu et al., 2014\)](#page-22-16), Gaussian Markov random field horseshoe prior [\(Denis and Tadesse, 2023\)](#page-22-17))

Results: Improvement in prediction quality depends on several factors such as quality of information, relevance to the trait considered, etc. [\(Peterson et al., 2016;](#page-22-10) [Mollandin](#page-22-18) [et al., 2022\)](#page-22-18)

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Idea: To use gene expression data or prior knowledge information into GS models

$$
Y = \mu + \underbrace{X}_{\text{gene expression data}} \beta + \varepsilon
$$

GS models may be used BUT questions about the interest of using transciptomic data instead of or in addition of genetic data.

Low gain in using transcriptomic data in prediction/Results vary according to environments [Chateigner et al. \(2020\)](#page-22-19):

Questions:

- How to predict a phenotype measured at one time point given that gene expressions vary over tissues, time, and environments?
- Do we need to provide more stable information ? Via a common graph structure obtained across multiple co-expression networks?

"The problem of identifying predictors that are both relevant to a response variable of interest and functionally related to one another."

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- ● Various statistical and biological questions raised...
- **But the bibliography is not exhaustive.... there are certainly already responses** to our questions….
- But seems interesting for biologists, geneticists, and statisticians
- \leftrightarrow Master student for working on the first part with Bénédicte Favreau (Biologist, Cirad) on Eucalyptus
- \leftrightarrow To continue exchanging on those subjects...

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