Integrating GENomic prediction with GENe regulatory networks to optimize genetic value prediction : biological and statistical challenges

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IGEN: A 2 days seminar in December 2022



- Biologists
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Need to infer gene expression network/graph for addressing two biological objectives:

- To gain insights into complex biological mechanisms involved in important processes, such as disease progress or growth
- To improve prediction of important phenotypes in genetic improvement context

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Towards a better understanding

- Network inference
- Network evaluation

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Towards a better understanding (genomic context)

To infer links/connections between genes for identifying biological mechanisms (such as key genes, functional modules, relations between network and a phenotype of interest, etc.)

Example:

How potassium and sodium fertilization impact biological mechanisms involved in response to water deficiency in *Eucalyptus grandis*?



Figure 1: Gene co-expression network (on left), bar plot representing the average gene significance of the genes within the cluster purple (middle), and the associated enrichment map (on right) (Favreau et al., 2019).

Network inference

How to build co-expression network from gene expression data?

Data:

$$X = \begin{pmatrix} X_{1,1} & X_{1,2} & \cdots & X_{1,n} \\ X_{2,1} & X_{2,2} & \cdots & X_{2,n} \\ \vdots & \vdots & \ddots & \vdots \\ X_{m,1} & X_{m,2} & \cdots & X_{m,n} \end{pmatrix}$$

with $X_{i,i}$ the expression level of gene *j* for sample *i*

We want to infer **network/graph** where:

- Vertices: genes
- Edges: links between genes (gene-gene interactions)

Network inference

What do we mean by links (gene-gene interactions)?

- Does it depend on biological question and/or experimental design?
- Does co-expression network aim at focusing on direct co-expression between ۲ genes? (Villa-Vialaneix et al., 2013; Grimes et al., 2019)

In the litterature

- Pearson-based correlation networks (relevance networks): marginal relationships between genes. Each pair of genes is considered alone: very dense networks, edges represent marginal connections not direct or causal
- Partial correlation based networks: direct relationships between genes. Correlation between two genes corrected for all other genes under investigation

Network inference



Gene2 and Gene3 correlated but not dependent on each other

Gaussian Graphical Models (GGMs) (Lauritzen, 1996) commonly used to estimate partial correlations

- Improve measurement of direct relations between gene expressions by accounting for the effect of all expression data
- More efficient for grouping together genes with a common function / more consistent to prior biological knowledge (Werhli et al., 2006; Krumsiek et al., 2011; Villa-Vialaneix et al., 2013)

Network inference: Gaussian Graphical Model

Let X_i be the *m*-vector (gene expression) of observed data for subject *i* such that

$$X_i \sim \mathcal{N}_m(\mu, \Sigma), \ i = 1, \ldots n,$$

with $\mu \in \mathbb{R}^m$ is the mean vector, Σ is the covariance matrix which is a positive semi-definite symmetric matrix, and $\Omega = \Sigma^{-1} \in \mathbb{R}^m \times \mathbb{R}^m$ is the precision matrix.

 \hookrightarrow Conditional independence implied by the form/structure of the precision matrix:

Gene j and Gene k are linked $\Leftrightarrow \Omega_{jk} > 0$.

Problem: When n < m, Σ is not full rank \Rightarrow can not be inverted

Network inference: Gaussian Graphical Model

Various estimation techniques (from the review done by Altenbuchinger et al. (2020)):

Nethod name	Software name	Reference	Parameter estimation	Model selection	Features	Availability
(c) Constian Oraphical Mode Graphical Lasso	k zława	1321	11 penalized maximum likelihood	-	Computationally efficient and sparse	R peckage https://CRAN.B-orelect.org/peckage=slasso
	0GMbdect	(81)	inference of laverse covariance matrix 6 different methods: CO1 [552] ; nol- wise regression [263] ; adopted II penalog [53] ; combination of CO3 and nole-wise regression; combination of CO3, node-wise regression; and adaptive II penalog; quais eshanative combination at neighborhood selection with different parameter combination rules	Minimization of penalized empirical risk (34)	solution Selection of penalisation parameters(s) of any graph estimation procedure and comparison of any collection of estimation procedures possible	R package https://CRAN.ik.project.org/package = 60Modect
Sparse Partial Correlation Estimation	space	[29]	Joint sparse regression model to simultaneously perform neighborhood selection for all nodes	NC-type criterion [29]	Method specifically designed for $p \gg N$ scenario, particularly powerful for hab- identification	R package https://CRAN.R.project.org/package=space
	diamly.	[85]	Graphical LASSO	EBIC or local FDR	Allows estimation of GGMs, graph visualization and analysis	R package https://CRAN.R project.org/package-ograph
High-Dimensional Undirected Graph Estimation	NUGE	[56]	Neighborhood selection (26) or graphical LASSO, further acceleration by lossy screening rule preselecting neighborhood of each node via thresholding sample correlation	STARS [36] , RIC, or EBIC for place	Integrates data preprocessing, neighborhood screening, graph estimation, and model selection techniques into one pipeline	R package https://CRAN.R-project.org/package=huge
Covariance Shrinkage	GeneNet	[25]	Analytic shrinkage estimation of covariance and (partial) correlation matrices	Parameter calibration according to [41] and significance thresholding using the local FDR	Very efficient, no parameter tuning, also suitable for dynamic (partial) correlations [57]	R package https://CRAN.R-project.org/package=GeneNet
	1960	[58]	Neighborhood selection [26] for GGMs	Stability selection [37] and STARS [36]	Allows estimation of GGMs, bing models, and Poisson family graphical models	R package https://CRAN.R-project.org/package=XMRF
	FeelOUM	[99]	ANT algorithm [33]		Efficient, taming free OGM estimation for large variable sets, supplies p-values and coefficience intervals for estimated edges	R package http://www.pitt.edu/~wee/f7/iist05M.html
	SLOGM	[60]	4 different methods: ANT algorithm [33], desparalified modewise socied LASSO [51], de separalified graphical LASSO [52], and (scaled) LASSO GGM semination with FER control [53]	FDR multiple testing	Provides confidence intervals, p-scores, and p-subses for estimated edges, faster than PossiCM	R package https://CRAN.B.project.org/package=SELGGM
	GeNeCK	[64]	Neighborhood selection, GeneNer, spece, glasse, glasso SF [65], Rayesian: glasse [66], SSPACE, and EGLASSO for GGMs	p-Value thresholding for ensemble-based network aggregation method [67]	Ensemble-based network aggregation method [67] allows combination of aetworks reconstructed by different methods	Web server http://ionbishpc.swmod.edu/geneck/

- More or less adapted for dealing with high-dimensional data: low to high differences observed
- More or less user friendly
- \Rightarrow Need guidelines for choosing the most adapted/to compare them

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Network evaluation

Statistical questions

Network evaluation

As most of co-expression networks in plants are Pearson-based correlation networks \Rightarrow Need to compare Pearson-based correlation network and partial correlation based network (Werhli et al., 2006; Krumsiek et al., 2011)

How to compare the inferred networks? How to evaluate their biological relevance?

- Functional enrichment analysis for testing the biological relevance, detection of key genes, relevance of networks to the phenotype of interest, etc. (Villa-Vialaneix et al., 2013; Lee et al., 2020)
- To compare to a "reference" network (obtained from data base such as STRING protein-protein interactions database)
- \hookrightarrow Which statistical measures?
 - Co-expression Differential Network Analysis: to extract the common structure (Grimes et al., 2019; Peterson et al., 2020)

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Is \ll functional understanding \gg relevant for prediction objectives, if it is the case how we take it into consideration ?

Idea: To use gene expression data or prior knowledge information into GS models

Genomic Selection (GS) model

$$Y = \mu + \underbrace{X\beta}_{GEBV} + \varepsilon$$

with $Y \in n \times 1$ the phenotype of interest, $X \in n \times p$ the marker matrix, $\beta \in p \times 1$ the marker effects, and $\varepsilon \sim \mathcal{N}_n(0, \sigma^2 Idn)$. GEBV: Genomic Estimated Breeding Value

 \hookrightarrow Various statistical approaches for estimating marker effects $\hat{\beta}$ (Ridge regression, Bayesian Lasso (BayesB), BayesC, etc)



Idea: To use gene expression data or **prior knowledge information** into GS models

$$Y = \mu + X \underbrace{\beta}_{\text{prior knowledge information}} + \varepsilon$$

Which type of information?

- From previous experimental studies (Co-expression networks, GO terms, GWAS results, selection signature,...) but may be not adequate with data at hand
- From "physical" knowledge: markers belonging to the same gene

Idea: To use gene expression data or **prior knowledge information** into GS models

$$Y = \mu + X \underbrace{\beta}_{\text{prior knowledge information}} + \varepsilon$$

"Although there are many databases that provide information on biochemical relationships under normal conditions, the available reference networks may be incomplete or inappropriate for the experimental condition or set of subjects under study" (Peterson et al., 2016)

- ↔ Need to use statistical approaches integrating different degrees of fidelity/belief to the prior knowledge (to guard against mis-specification) (Stingo et al., 2010; Kundu et al., 2018; Denis et al., 2022)
- \hookrightarrow Need to use statistical approaches providing a trade-off between prior knowledge and computational complexity

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Idea: To use gene expression data or **prior knowledge information** into GS models



How to integrate those information into GS models?

Bayesian framework is a natural framework where prior knowledge may be specified via prior on regression coefficients (Bayesian fused and group Lasso (Kyung et al., 2010), Ising prior (Li and Zhang, 2010)) *Example:* $\beta \sim \mathcal{N}_p(0, \Sigma)$ with Σ related to structure between variables specified via for instance by undirected graph (Graph Laplacian prior(Liu et al., 2014), Gaussian Markov random field horseshoe prior (Denis and Tadesse, 2023))

Results: Improvement in prediction quality depends on several factors such as quality of information, relevance to the trait considered, etc. (Peterson et al., 2016; Mollandin et al., 2022)

Idea: To use gene expression data or prior knowledge information into GS models

$$Y = \mu + \underbrace{X}_{\text{gene expression data}} \beta + \varepsilon$$

GS models may be used BUT questions about the interest of using transciptomic data instead of or in addition of genetic data.

Low gain in using transcriptomic data in prediction/Results vary according to environments Chateigner et al. (2020):

Questions:

- How to predict a phenotype measured at one time point given that gene expressions vary over tissues, time, and environments?
- Do we need to provide more stable information ? Via a common graph structure obtained across multiple co-expression networks?

"The problem of identifying predictors that are both relevant to a response variable of interest and functionally related to one another."

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- Various statistical and biological questions raised...
- But the bibliography is not exhaustive.... there are certainly already responses to our questions....
- But seems interesting for biologists, geneticists, and statisticians
- ↔ Master student for working on the first part with Bénédicte Favreau (Biologist, Cirad) on Eucalyptus
- $\, \hookrightarrow \,$ To continue exchanging on those subjects...

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Bibliography

- Altenbuchinger, M., Weihs, A., Quackenbush, J., Grabe, H. J., and Zacharias, H. U. (2020). Gaussian and mixed graphical models as (multi-) omics data analysis tools. Biochimica et Biophysica Acta (BBA)-Gene Regulatory Mechanisms, 1863(6):194418.
- Chateigner, A., Lesage-Descauses, M.-C., Rogier, O., Jorge, V., Leplé, J.-C., Brunaud, V., Roux, C. P.-L., Soubigou-Taconnat, L., Martin-Magniette, M.-L., Sanchez, L., et al. (2020). Gene expression predictions and networks in natural populations supports the omnigenic theory. BMC genomics, 21(1):1–16.

Denis, M. and Tadesse, M. G. (2023). Graph-structured variable selection with gaussian markov random field horseshoe prior. HAL.

- Denis, M., Varghese, R. S., Barefoot, M. E., Tadesse, M. G., and Ressom, H. W. (2022). A bayesian two-step integrative procedure incorporating prior knowledge for the identification of mirna-mrnas involved in hepatocellular carcinoma. In 2022 44th Annual International Conference of the IEEE Engineering in Medicine & Biology Society (EMBC), pages 81–86. IEEE.
- Favreau, B., Denis, M., Ployet, R., Mounet, F., Peireira da Silva, H., Franceschini, L., Laclau, J.-P., Labate, C., and Carrer, H. (2019). Distinct leaf transcriptomic response of water deficient eucalyptus grandis submitted to potassium and sodium fertilization. PLoS One, 14(6):e0218528.

Grimes, T., Potter, S. S., and Datta, S. (2019). Integrating gene regulatory pathways into differential network analysis of gene expression data. Scientific reports, 9(1):5479.

- Krumsiek, J., Suhre, K., Illig, T., Adamski, J., and Theis, F. J. (2011). Gaussian graphical modeling reconstructs pathway reactions from high-throughput metabolomics data. BMC systems biology, 5:1-16.
- Kundu, S., Cheng, Y., Shin, M., Manyam, G., Mallick, B. K., and Baladandayuthapani, V. (2018). Bayesian variable selection with graphical structure learning: Applications in integrative genomics. *PloS one*, 13(7):e0195070.
- Kyung, M., Gill, J., Ghosh, M., Casella, G., et al. (2010). Penalized regression, standard errors, and Bayesian lassos. Bayesian Analysis, 5(2):369-411.

Lauritzen, S. L. (1996). Graphical models, volume 17. Clarendon Press.

- Lee, J., Shah, M., Ballouz, S., Crow, M., and Gillis, J. (2020). Cococonet: conserved and comparative co-expression across a diverse set of species. Nucleic acids research, 48(W1):W566–W571.
- Li, F. and Zhang, N. R. (2010). Bayesian variable selection in structured high-dimensional covariate spaces with applications in genomics. Journal of the American Statistical Association, 105(491):1202–1214.
- Liu, F., Chakraborty, S., Li, F., Liu, Y., Lozano, A. C., et al. (2014). Bayesian regularization via graph Laplacian. Bayesian Analysis, 9(2):449-474.
- Mollandin, F., Gilbert, H., Croiseau, P., and Rau, A. (2022). Accounting for overlapping annotations in genomic prediction models of complex traits. BMC bioinformatics, 23(1):1-22.
- Peterson, C. B., Osborne, N., Stingo, F. C., Bourgeat, P., Doecke, J. D., and Vannucci, M. (2020). Bayesian modeling of multiple structural connectivity networks during the progression of alzheimer's disease. *Biometrics*, 76(4):1120–1132.
- Peterson, C. B., Stingo, F. C., and Vannucci, M. (2016). Joint bayesian variable and graph selection for regression models with network-structured predictors. Statistics in medicine, 35(7):1017–1031.
- Stingo, F. C., Chen, Y. A., Vannucci, M., Barrier, M., and Mirkes, P. E. (2010). A bayesian graphical modeling approach to microrna regulatory network inference. The annals of applied statistics, 4(4):2024.
- Villa-Vialaneix, N., Liaubet, L., Laurent, T., Cherel, P., Gamot, A., and SanCristobal, M. (2013). The structure of a gene co-expression network reveals biological functions underlying eqtls. PloS one, 8(4):e60045.
- Werhli, A. V., Grzegorczyk, M., and Husmeier, D. (2006). Comparative evaluation of reverse engineering gene regulatory networks with relevance networks, graphical gaussian models and bayesian networks. Bioinformatics, 22(20):2523-2531.