

Automated generation of resource allocation models at cellular scales

from prokaryotic to eukaryotic cells

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Resource allocation as a strong design principle in living organisms

Organisms



Models based on resource allocation

Biomass allocation by source-sink empirical relations

Nutrient partitioning model between life stages

- ➔ High prediction capability despite their « simplicity »
- ➔ Resource allocation between organs and life functions is a strong structural constraint



At cellular scale?

Resource allocation in bacteria

MICROBIOLOGICAL REVIEWS, June 1991, p. 316-333
0146-0749/91/020316-18\$02.00/0
Copyright © 1991, American Society for Microbiology

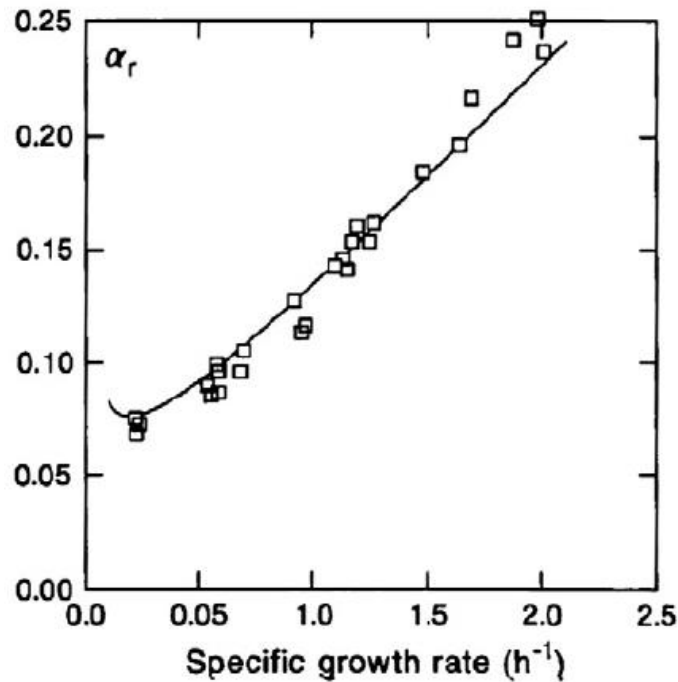
Vol. 55, No. 2

Growth Rate of *Escherichia coli*

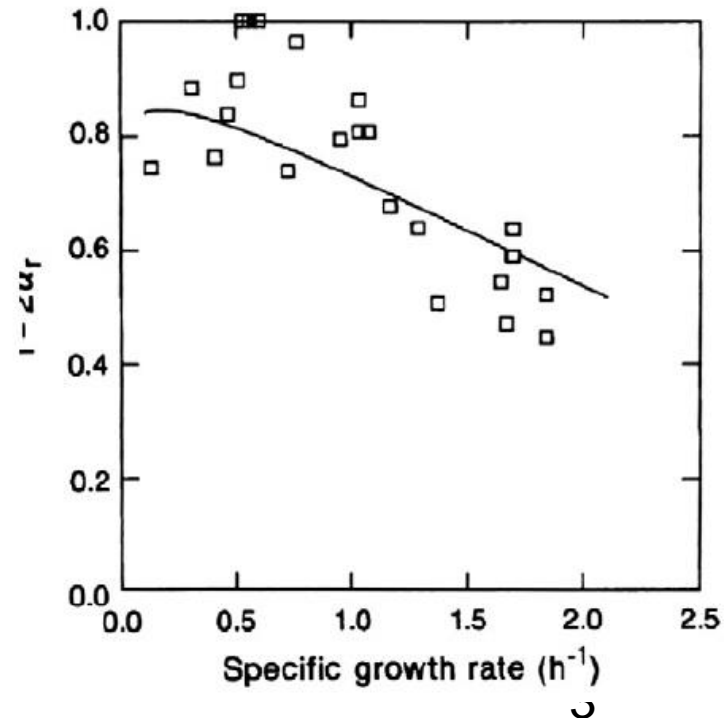
ALLEN G. MARR

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Ribosomes



Non-ribosomal proteins



A new perspective since 2009

How to predict resource allocation at genome scale?

PERSPECTIVE (2009)

**Shifts in growth strategies reflect tradeoffs
in cellular economics**

**Nonlinear optimization
problem**

Douwe Molenaar^{1,3,4,5,*}, Rogier van Berlo^{2,4}, Dick de Ridder^{2,4}
and Bas Teusink^{1,3,4,5}

**Interdependence of Cell Growth
and Gene Expression: (2010)
Origins and Consequences**

Not at genome-scale W. Gunderson,^{2*} Eduard M. Mateescu,¹ Zhongge Zhang,² Terence Hwa^{1,2,‡}

Joint 48th IEEE Conference on Decision and Control and
28th Chinese Control Conference
Shanghai, P.R. China, December 16-18, 2009

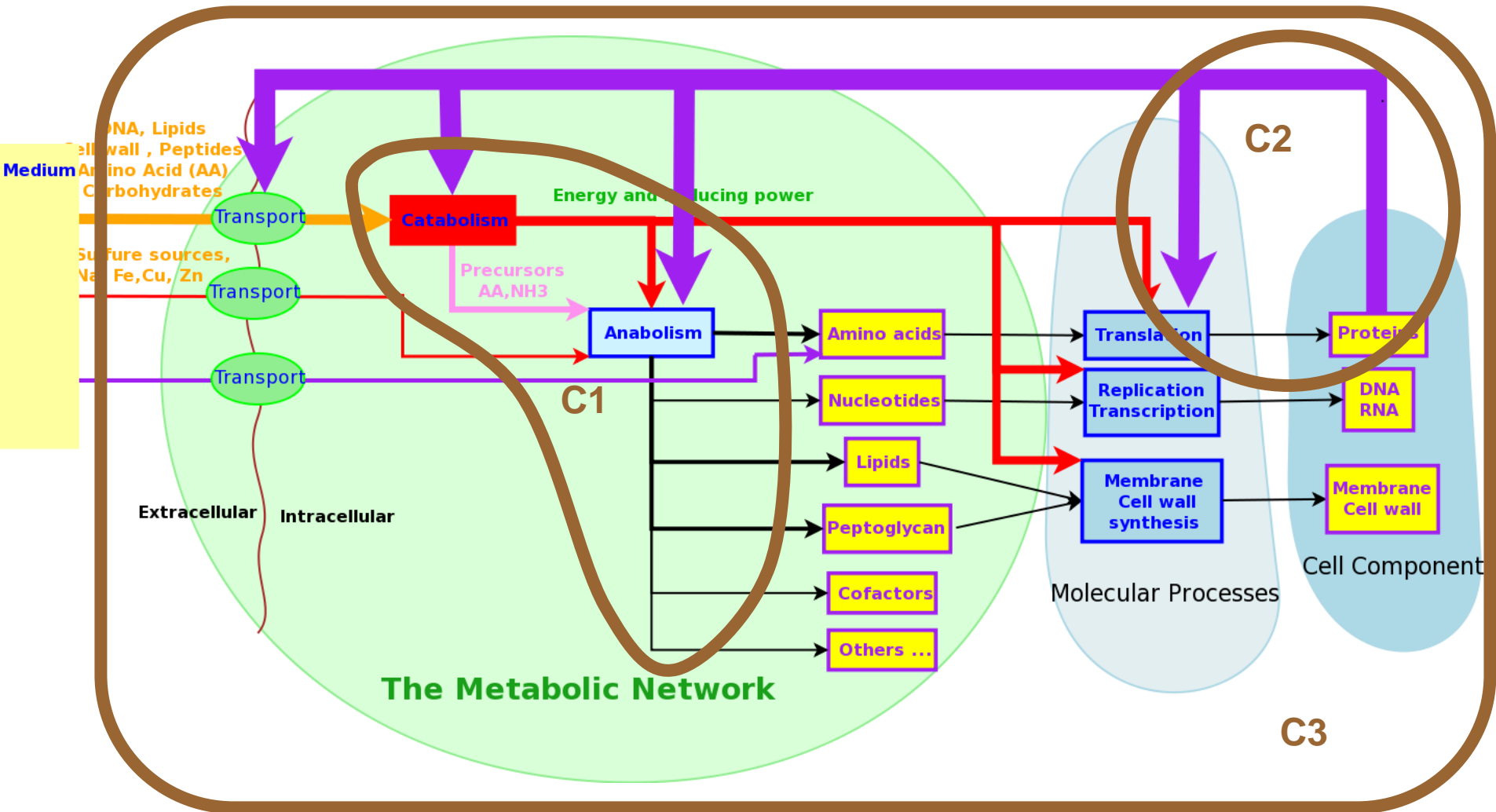
(2009)

Convex and genome-scale

Cell design in bacteria as a convex optimization problem

Anne Goelzer, Vincent Fromion and Gérard Scorletti

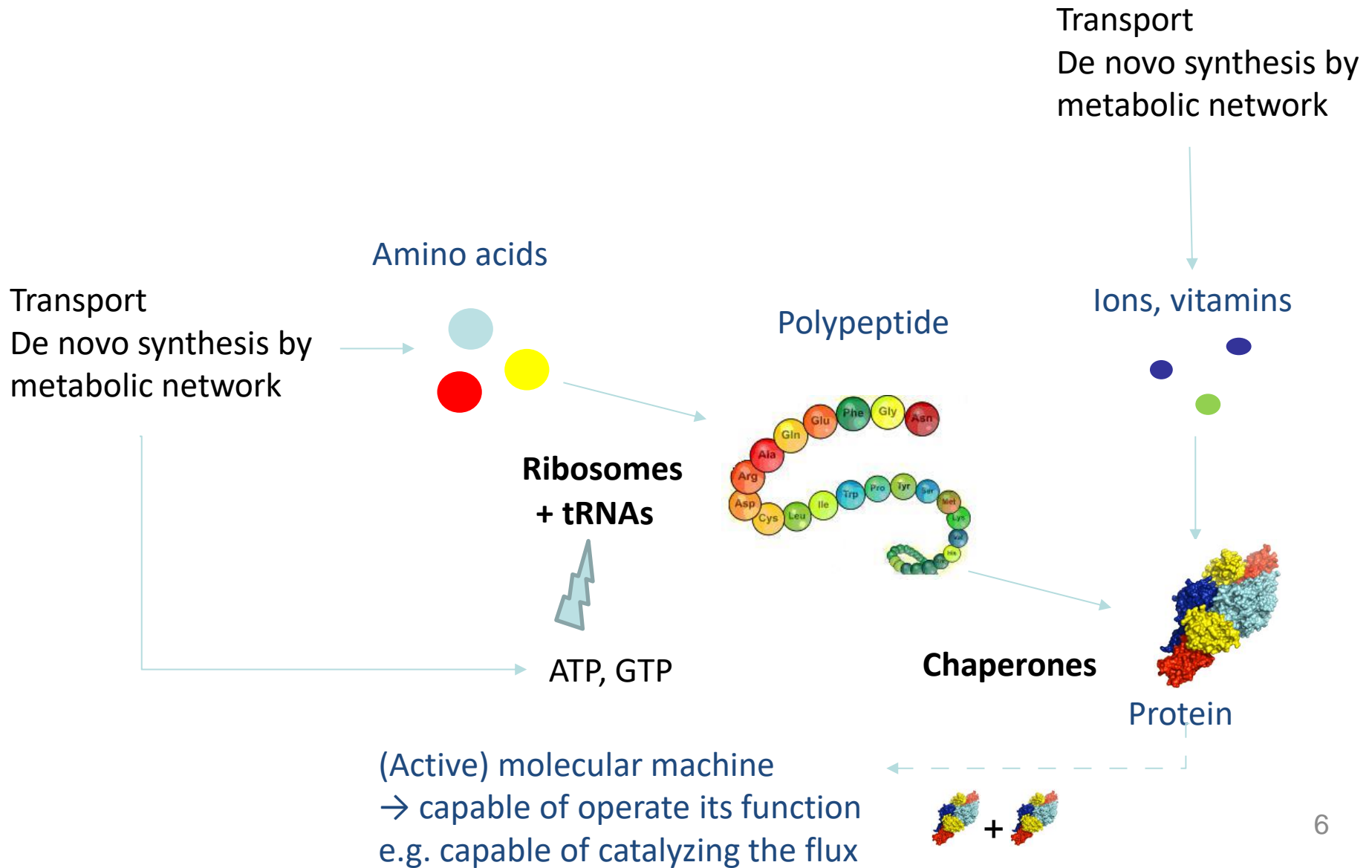
Three (main) structural constraints



Resources (especially proteins) have to be shared by all biological processes (implicit feedback).

Resource sharing imposes constraints on cellular processes

Detailed integration of production costs for protein synthesis



Formalization into an optimization problem

Resource Balance Analysis (RBA)

For fixed $P_G \geq 0, \mu \geq 0$,

Find $R \geq 0, C \geq 0, \nu^x \in \mathcal{R}^m, |\nu_i| \leq k_{E_i} E_i$
 subject to

Energy & precursors
production

(C_{1a}) For all $i \in I_p$,

$$-\sum_{j=1}^m S_{p_{ij}} \nu_j^x + \mu \left(\sum_{j=1}^m C_{M_{ij}}^{M_p} |\nu_j^x| + C_{R_i}^{M_p} R + C_{C_i}^{M_p} C + C_{G_i}^{M_p} P_G^{x,T} \right) - \nu_Y = 0$$

(C_{1b}) For all $i \in I_c$,

$$-\sum_{j=1}^m S_{c_{ij}} \nu_j^x + \mu \bar{X}_{c_i} = 0$$

(C_{1c}) For all $i \in I_r$,

$$\sum_{j=1}^m S_{r_{ij}} \nu_j^x + \mu \left(\sum_{j=1}^m C_{M_{ij}}^{M_r} |\nu_j^x| + C_{R_i}^{M_r} R + C_{C_i}^{M_r} C + C_{G_i}^{M_r} P_G^{x,T} \right) = 0$$

(C_{1d}) For all $i \in I_i$,

$$\sum_{j=1}^m S_{I_{ij}} \nu_j^x = 0$$

(C_{2a})
$$\mu \left(\sum_{j=1}^m C_{M_j}^R |\nu_j^x| + C_R^R R + C_C^R C + C_G^R P_G^{x,T} \right) - k_T R = 0$$

Protein production

(C_{2b})
$$\alpha_c \mu \left(\sum_{j=1}^m C_{M_j}^R |\nu_j^x| + C_R^R R + C_C^R C + C_G^R P_G^{x,T} \right) - k_C C = 0$$

Protein folding

(C_{3a})
$$\sum_{j=1}^m C_{M_j}^D |\nu_j^c| + C_R^D R + C_C^D C + C_G^D P_G^{c,T} - \bar{D}_c \leq 0$$

Cytosol occupancy

(C_{3b})
$$\sum_{j=1}^m C_{M_j}^S |\nu_j^s| + C_G^S P_G^{s,T} - \bar{D}_s \leq 0$$

Membrane occupancy

A. Goelzer, V. Fromion and G. Scorletti *Cell design in bacteria as a convex optimization problem*. 48th IEEE Conference on Decision and Control, China, 4517 -22. **2009**.

A. Goelzer, V. Fromion and G. Scorletti *Cell design in bacteria as a convex optimization problem*. Automatica,47(6):1210-1218. **2011**.

The RBA framework

- ❑ The feasibility problem is convex
- ❑ Equivalence with a Linear Programming (LP) optimization problem
 - ➡ same complexity as FBA, efficient resolution at genome scale!

- ❑ For a set of **given extracellular nutrient concentrations**, we can prove that there exists a **maximal growth rate value**
 - **without setting an objective function** (contrary to FBA);
 - **defined by a trade-off on the resource allocation** (especially on proteins);
 - for which a resource distribution (enzyme/ribosomes) exists;
 - and can be efficiently computed through the iterative resolution of LP optimization problems;

- ❑ Every mechanism saving resources increases the growth rate

- ❑ Theoretical prediction of induced/repressed sub-systems in the metabolic network (towards the prediction of genetic regulations)

- ➡ RBA computes (a) the maximal **growth rate**, (b) the **metabolic fluxes** including the **substrate uptake and by-product secretion rates**, and (c) the genome-scale resource allocation including the **absolute abundances of enzymes, transporters, ribosomes, and chaperones**, i.e. the **phenotype** of the organism

Rewriting the RBA problem in a more compact way

For fixed $P_G \geq 0$, $\mu \geq 0$,

find $Y \in \mathbb{R}_{\geq 0}^{m+p}$, $\nu \in \mathbb{R}^m$,

subject to

$$(C_1) \quad -\Omega\nu + \mu(C_Y^S Y + C_B^S \bar{B} + C_G^S P_G) = 0$$

$$(C_{2a}) \quad \mu(C_Y^M Y + C_G^M P_G) - K_T Y \leq 0$$

$$(C_{2b}) \quad -K_E' Y \leq \nu \leq K_E Y$$

$$(C_3) \quad C_Y^D Y + C_G^D P_G - \bar{D} \leq 0$$

752 parameters to be estimated

For fixed $P_G \geq 0$, $\mu \geq 0$,

find $Y \in \mathbb{R}_{\geq 0}^{m+p}$, $\nu \in \mathbb{R}^m$,

subject to

$$(C_1) \quad -\Omega\nu + \mu(C_Y^S Y + C_B^S \bar{B} + C_G^S P_G) = 0$$

$$(C_{2a}) \quad \mu(C_Y^M Y + C_G^M P_G) - K_T Y \leq 0$$

$$(C_{2b}) \quad -K_E' Y \leq \nu \leq K_E Y$$

$$(C_3) \quad C_Y^D Y + C_G^D P_G - \bar{D} \leq 0$$

From the stoichiometry
of chemical reactions

From annotation
& bioinformatics

752 parameters to be estimated

Physiology

For fixed $P_G \geq 0, \mu \geq 0,$

Proteomics
(Q., RA.)

Proteomics
(Q., RA.)

Fluxomics

From literature
or biomass comp
or data.

find

$$Y \in \mathbb{R}_{\geq 0}^{m+p}, \nu \in \mathbb{R}^m,$$

subject to

$$(C_1) \quad -\Omega\nu + \mu(C_Y^S Y + C_B^S \bar{B} + C_G^S P_G) = 0$$

$$(C_{2a}) \quad \mu(C_Y^M Y + C_G^M P_G) - K_T Y \leq 0$$

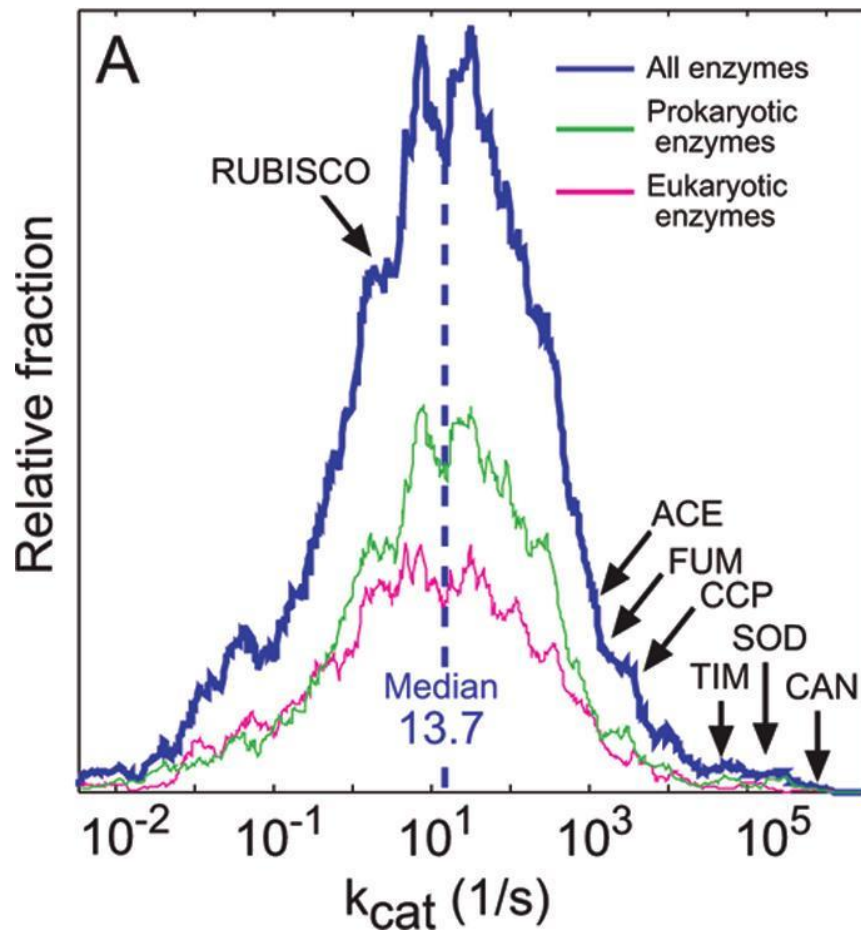
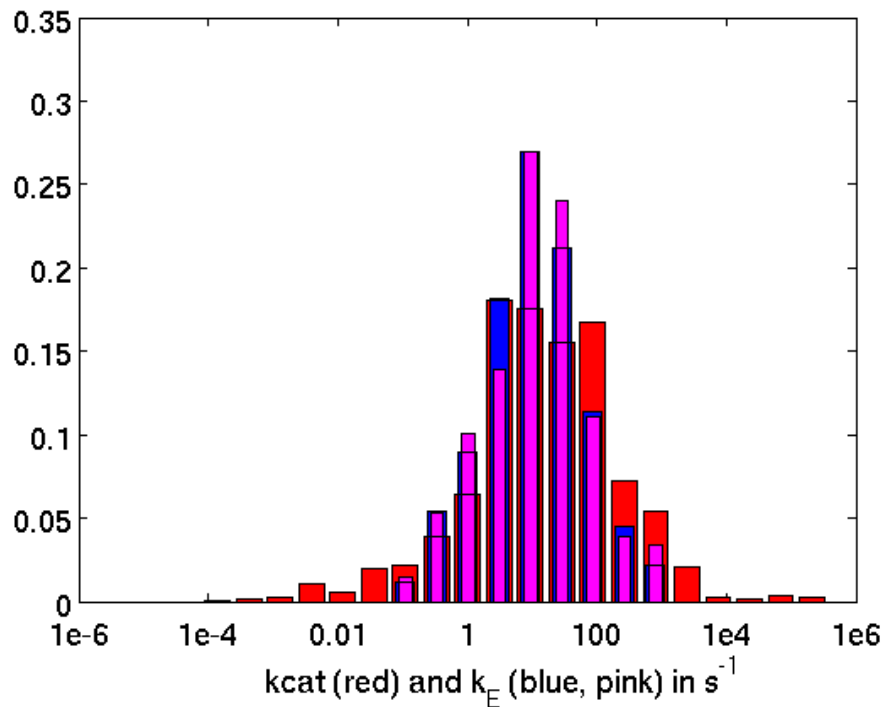
$$(C_{2b}) \quad -K_E Y \leq \nu \leq K_E Y$$

$$(C_3) \quad C_Y^D Y + C_G^D P_G - \bar{D} \leq 0$$

Q.: quantitative
RA.: relative/absolute

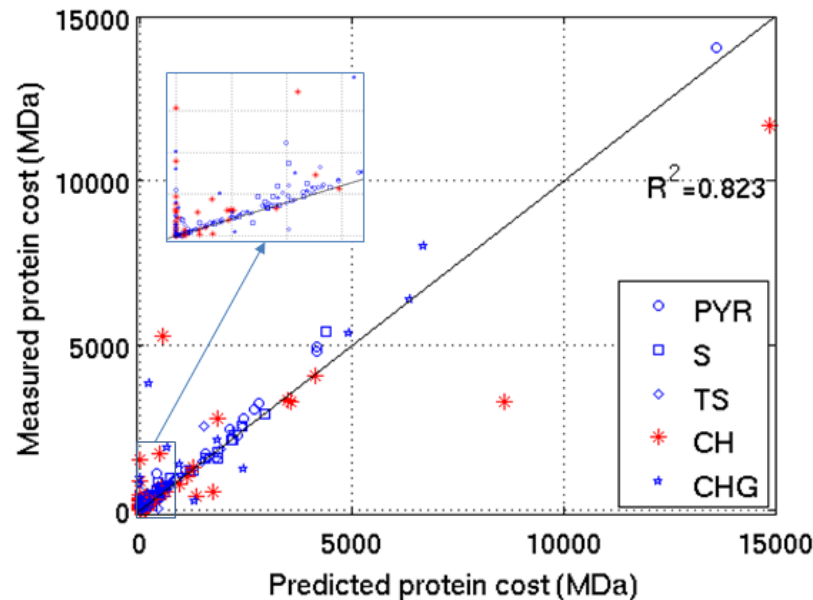
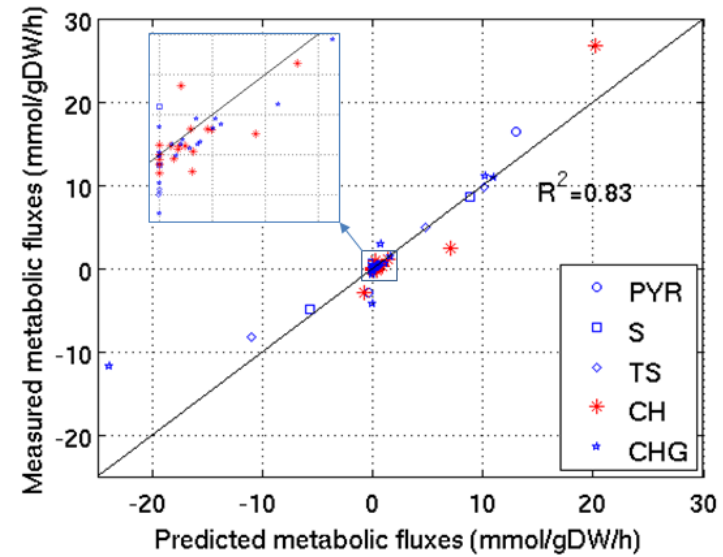
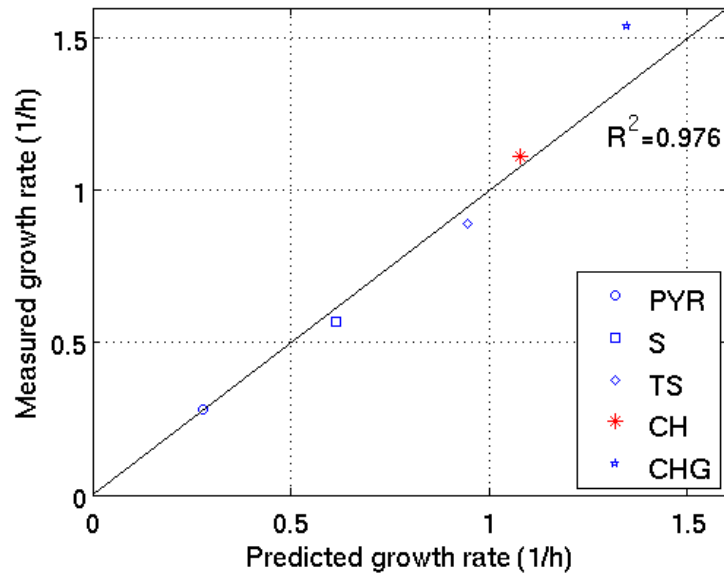
Total protein content + proteomics + protein localization

Identification of apparent catalytic rate of ≈ 600 enzymes (Consistency with the expected distribution)



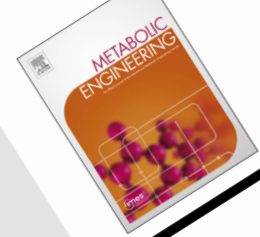
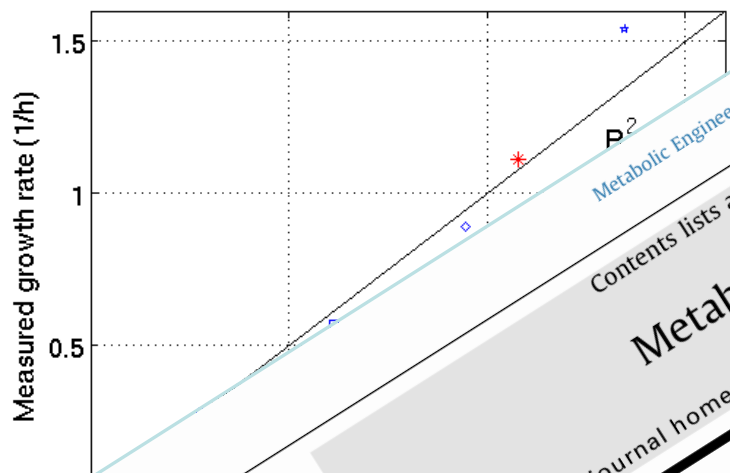
A. Bar-Even, et al. The Moderately Efficient Enzyme: Evolutionary and Physicochemical Trends Shaping Enzyme Parameters, *Biochemistry*, 2011, 50 (21), pp. 4402–4410

Quantitative prediction of the resource allocation between 72 cellular processes



Quantitative prediction of the resource allocation in cellular growth

between 72



Metabolic Engineering 32 (2015) 232–243

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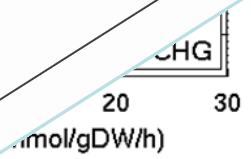
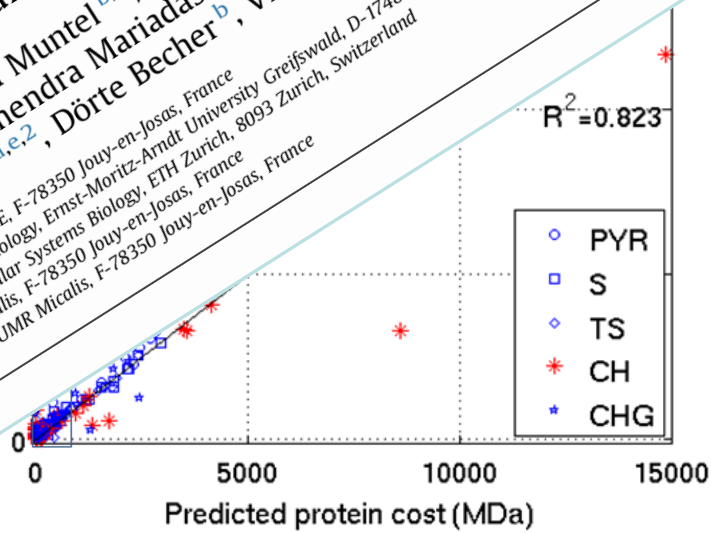


Original Research Article

Quantitative prediction of genome-wide resource allocation in bacteria

Anne Goelzer^a, Jan Muntel^{b,1}, Victor Chubukov^c, Matthieu Jules^{d,e}, Eric Prestel^{d,e},
 Rolf Nölker^b, Mahendra Mariadassou^a, Stéphane Aymerich^{d,e}, Michael Hecker^b,
 Philippe Noirot^{d,e,2}, Dörte Becher^b, Vincent Fromion^{a,*}

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Generation of RBA models for prokaryotes

What do we need to create a RBA model?

A metabolic network including the description of **enzymatic complexes**

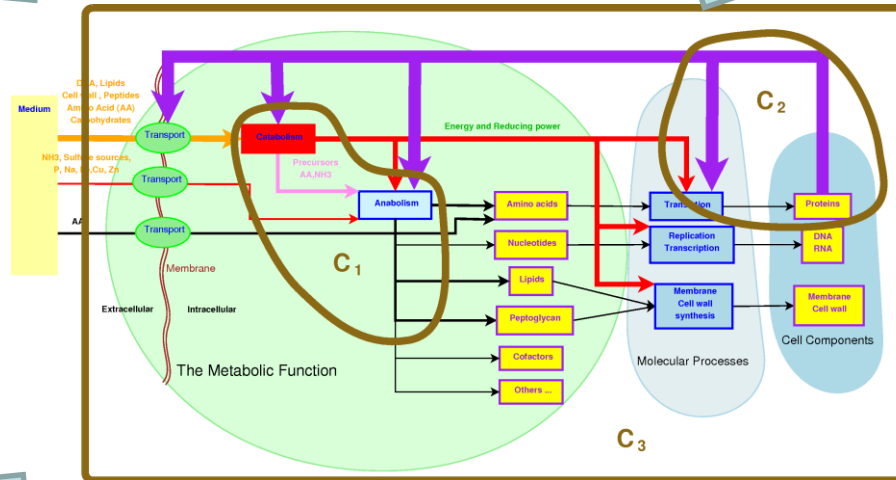
Description of non-metabolic molecular machines

Ribosomes, chaperones, etc.

Literature ou automatic reconstruction



Literature, Databases



Optimal resource allocation

Growth rate
Metabolic fluxes,
protein content

Uniprot

Information on protein sequences

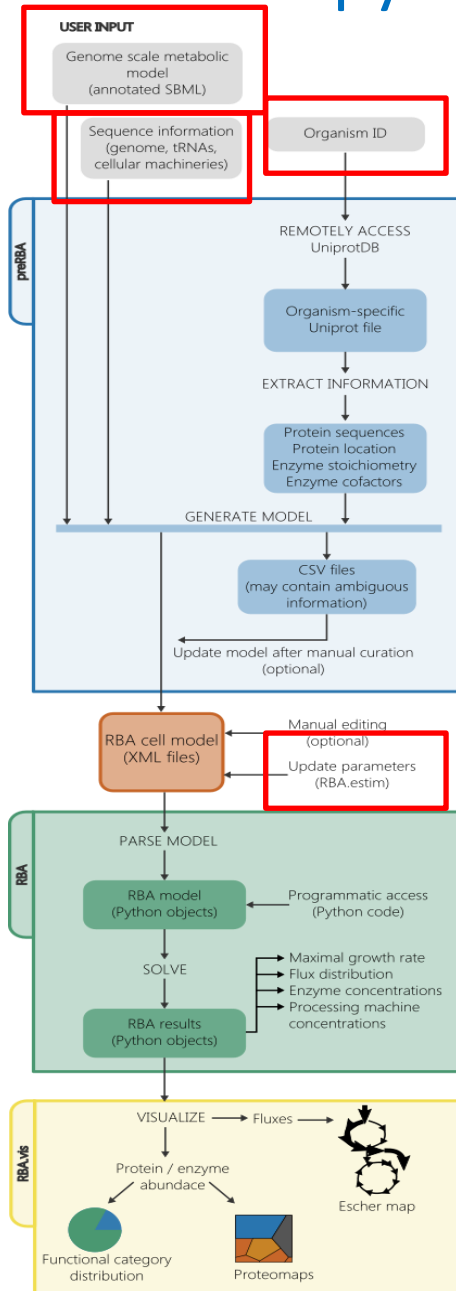
Amino acid sequences, Localization ions, cofactors, vitamins, structure, ...

Literature or Data

Parameters

Efficiencies of molecular machines, P_G amount, etc.

RBApy: a software system for bacterial resource allocation models



Inputs (mandatory):

- ❑ A genome-scale metabolic model (GSMM) including gene association (i.e. boolean AND/OR)
- ❑ The NCBI Taxon ID

Additional inputs (for model refinement)

- ❑ Definition of molecular machines,
- ❑ Composition of rRNAs, tRNAs

Additional inputs (for model calibration)

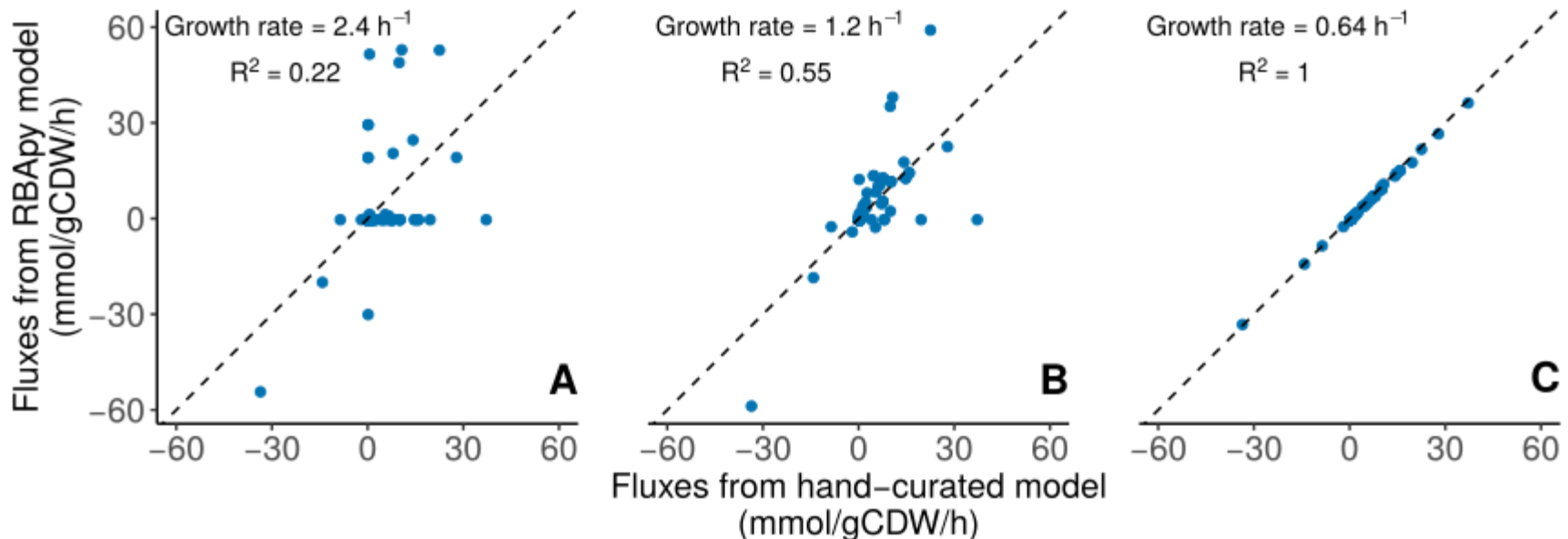
- ❑ Quantitative (or relative-absolute) proteomics
- ❑ Fluxomics (or prediction of metabolic fluxes)

Outputs:

- ❑ A RBA model in XML files
- ❑ Simulation results in text files

Validation of RBApy on *Bacillus subtilis*

- (A) Model created from SBML and Uniprot files with default parameters and default processes, where automatic merging for some identifiers failed (e.g. tRNAs IDs)
- (B) After matching SBML metabolite identifiers with Uniprot cofactor identifiers, and RBApy identifiers for metabolites involved in processes
- (C) After calibration of molecular machine efficiencies, adjustment of subunit stoichiometries of enzyme complexes and molecular machines from the hand-curated model, and adding metabolic demands for flagella movement and membrane biosynthesis.



A RBA model for *Escherichia coli* created from scratch

Source of information:

- ❑ The iJO1366 metabolic model [1]
- ❑ Quantitative proteomics [2] and fluxomics [3]
- ❑ Literature, Uniprot

Estimation of parameters:

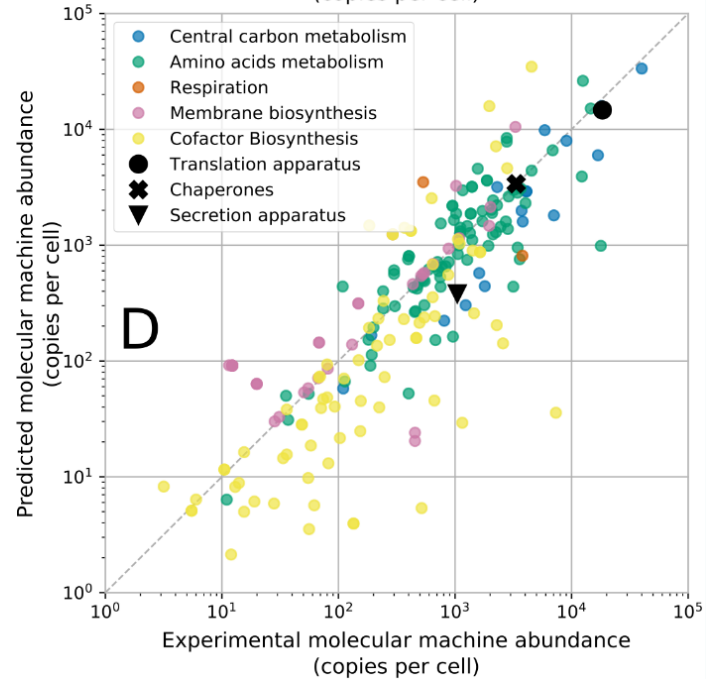
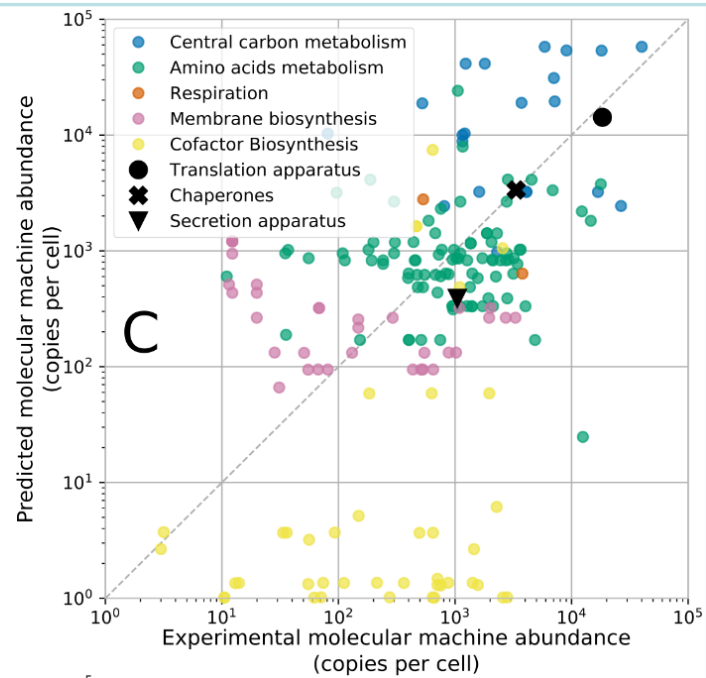
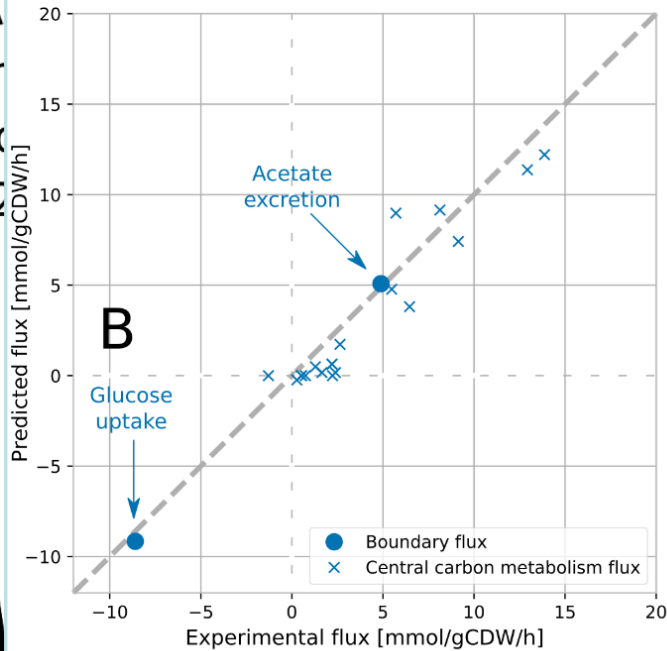
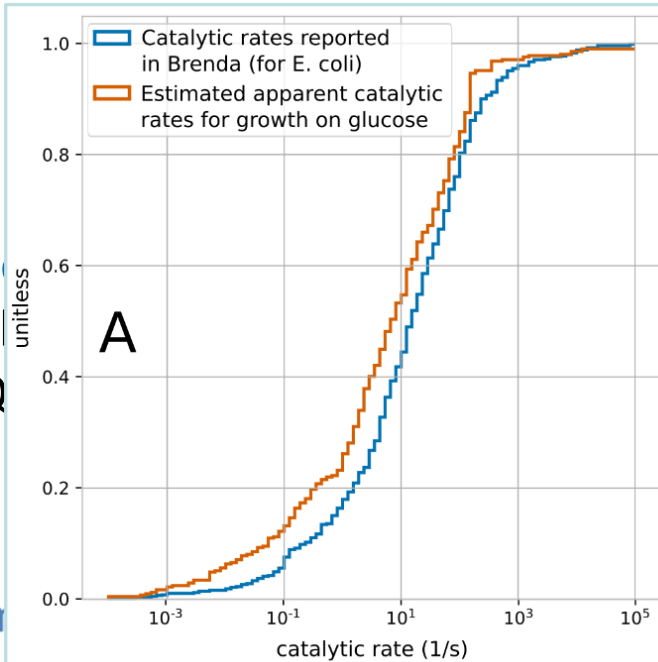
- ❑ Apparent catalytic rates k_E , efficiencies of molecular machines in glucose minimal medium (data from [2]+[3])
- ❑ Total protein abundance per compartment wrt growth rate, etc. based on [2]

Source

□ T
□ Q

Estim

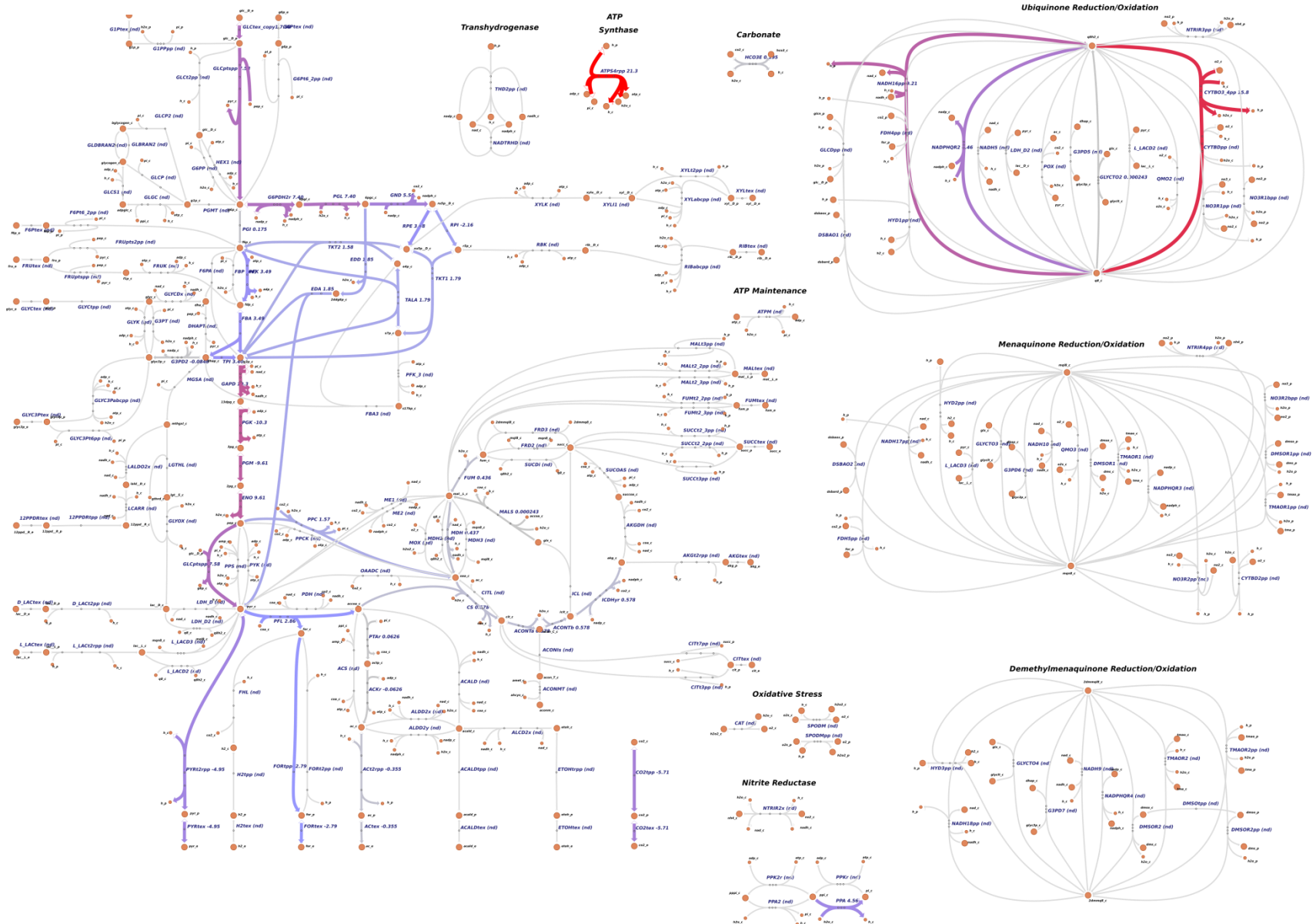
□ A
□ m
□ T
□ [2



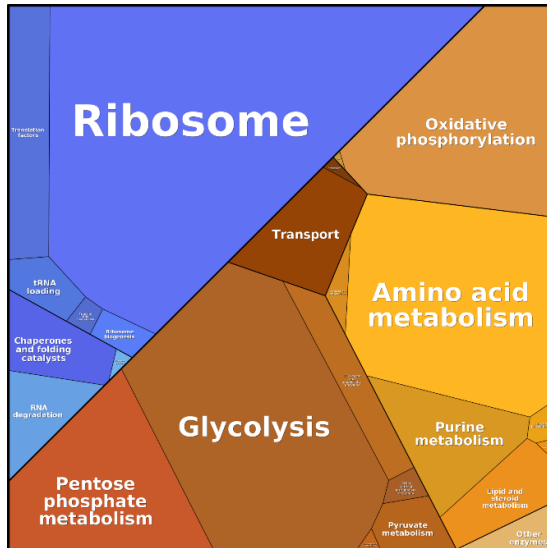
glucose
based on

IM

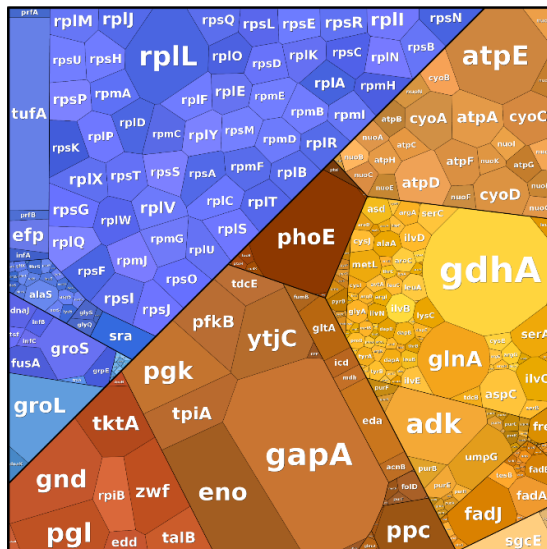
Flux visualization using Escher maps



Protein visualization using Proteomaps



Predicted protein abundances



Need additional information for the use of Proteomaps and Escher maps

- ❑ BIGG identifiers (Escher [1])
- ❑ Functional annotation (Proteomaps [2])

[1] King et al. Plos Comp. Biol. 2015, 11(8):e1004321

[2] Liebermeister et al. PNAS 2014, 111(23): 8488-8493

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Automated generation of bacterial resource allocation models

Ana Bulović^{b,1}, Stephan Fischer^{a,1}, Marc Dinh^a, Felipe Golib^a, Wolfram Liebermeister^{a,c}, Christian Poirier^a, Laurent Tournier^a, Edda Klipp^b, Vincent Fromion^{a,**}, Anne Goelzer^{a,*}

A versatile modeling framework

A new **cellular process** can be included straightforward by adding new capability constraints and new decision variables in RBA

- Necessitate to detail the production cost
- Introduce new parameters to identify

A new **compartment** can be included straightforward by adding new density constraints

- Introduce new parameters to identify

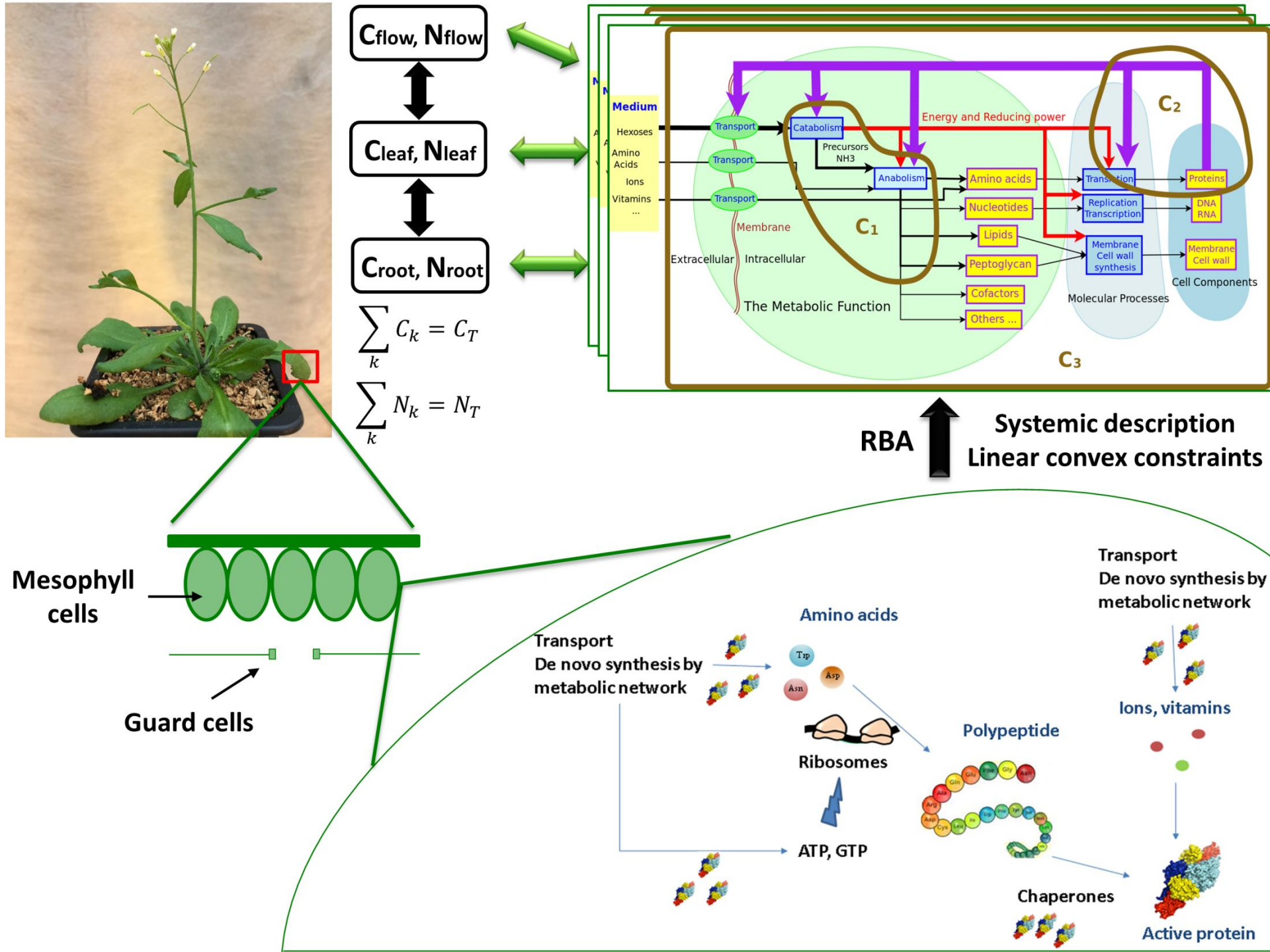


Possible currently by editing the XML files manually of RBApy



Future RBApy releases will contain tools to facilitate the integration of other processes and compartments

Towards RBA models for eukaryotic cells



Formalization into a LP optimization problem

$P_{rba}^e(\mu)$: For a fixed vector of concentrations $P_G \in \mathbb{R}_{>0}^{N_g}$, and the growth rate $\mu \geq 0$ of the cell,

find $Y \in \mathbb{R}_{\geq 0}^{N_y}, \nu \in \mathbb{R}^{N_m}, f \in \mathbb{R}_{\geq 0}^{N_c}$,

$$(C_1) \quad -\Omega\nu + \mu(C_Y^S Y + C_G^S P_G + C_B^S \bar{B} + C_F^S f \hat{B}) = 0$$

$$(C_2) \quad \mu(C_Y^M Y + C_G^M P_G) - K_T Y \leq 0$$

$$-K_E' Y \leq \nu \leq K_E Y$$

$$(C_{3a}) \quad C_Y^{D,iq} Y + C_G^{D,iq} P_G - C_F^{D,iq} f \leq 0$$

$$(C_{3b}) \quad C_Y^{D,eq} Y + C_G^{D,eq} P_G - C_F^{D,eq} f = 0$$

$$(C_{3c}) \quad C_F^F f - \bar{C} = 0$$

$$(C_{3d}) \quad \underline{f}_V \leq I_V f \leq \bar{f}_V$$

Capacity of
mol. machines

Constraints occupancy
on volume and membrane
compartments

Energy & precursors
production

- For fixed growth rate the optimization problem is a LP problem
- There exists a maximal growth rate μ^* such as $P_{rba}(\mu)$ is feasible for lower μ values, and unfeasible for upper values
- The optimal μ^* can be computed by a bisection algorithm by solving a series of LP.

Formalization into a LP problem

$P_{rba}^e(\mu)$: For a fixed vector μ

≥ 0 of the cell,

End

RBA for eukaryotic cells: foundations and theoretical developments

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Abstract

Resource allocation models were recently identified as new ways to investigate cell design principles. In particular, the Resource Balance Analysis (RBA) framework is the first constraint-based modelling method capable of accurate quantitative predictions of the genome-wide resource allocation. Initially developed and validated on bacteria, the objective of this paper is to provide the mathematical foundations of the extension of the RBA framework to eukaryotic cells. We especially investigate the way to handle the cellular compartments in order to formalize eventually the functioning of organelles. It turns out that the final RBA problem for eukaryotic cells is close to the one of prokaryotic cells. A theoretical point of view. The mathematical properties that were already identified from the prokaryotic RBA framework can be easily transposed to eukaryotic cells. In particular, the eukaryotic RBA problem can be solved easily at the cell scale by Linear Programming. This paves the way to future developments of RBA models for eukaryotic cells.

- For fixed growth rate μ
- There exists a μ^* such that $P_{rba}^e(\mu)$ is feasible for lower μ values, and unfeasible for $\mu > \mu^*$
- The optimal μ^* can be determined by a bisection algorithm by solving a series of LP.

What are the parameters ?

- K_E K'_E : the apparent catalytic rates of enzymes and transporters.
→ Literature/database or Total protein content, proteomics, fluxomics
- K_T : the efficiency of macromolecular machines as ribosomes.
→ Literature or Total protein content, proteomics, Ribosome abundance
- P_G : Abundance of proteins (per compartment) for which the activity is not explicitly described in the model.
→ Total protein content, proteomics, protein localization
- \bar{B} : Abundance of macrocomponents of biomass as total DNA, mRNA, cell wall, Lipids, Starch, free AA, etc.
→ Biomass composition
- \bar{C} : Link between compartments and membranes of organelles as the surface/volume ratio. → Literature



RBA models for prokaryotic and eukaryotic cells are highly similar !

Only a few changes to RBApy are necessary

- Improve the protein localization management
- Include additional cellular processes in compartments (e.g. translation process in mitochondrion and in cytoplasm) by default
- Implement the additional constraints related to compartment management
- Tools to facilitate the manual curation



A proof-of-concept on the leaf of *Arabidopsis thaliana* (*under progress*)

RBA for other eukaryotic cells (1/2)

« Straightforward » in theory, but ...

Problem in information description within GSMM ...

- ❑ Standards such as SBML need to evolve to account for molecular machine descriptions and template-based chemical reactions
- ❑ Need to unify identifiers of molecules, reactions, cellular functions across regna (plant, mammals, microorganisms)

... and beyond

- ❑ Molecular machines (including enzymatic complexes) are poorly described in databases (but recent progress like Reactome)
- ❑ Use of ontologies to formally describe the organism as a « system » (i.e. composed of molecular machines dedicated to a cellular function)
- ❑ Transfer of the knowledge from a model organism to another one (genericity vs specificity)

—————> Strong link with the knowledge representation field in biology

RBA for other eukaryotic cells (2/2)

Heterogeneity in organism description

Mammalian cells

- ❑ Active community to build a curated Human GSMM including gene association
- ❑ High degree of homology between mammals
- ❑ Automatic reconstruction based on orthologues search for common metabolic pathways
- ❑ Use of transcriptomics or proteomics to specialize GSMM per organ

—————→ Mouse GSMM generated from this procedure

Availability of data for RBA
parameters identification ?

Plant cells

- ❑ GSMMs usually do not include the description of enzymatic complexes (AND/OR rules)
- ❑ Difficulty to link genes Ids in GSMM and Uniprot
 - Lack of a unified standard between databases
- ❑ GSMMs need to be specialized by organs and by developmental stages
- ❑ Localization of isoforms could be improved by transcriptomics or proteomics

—————→ Strong link with the bioinformatics community to infer sequence-based information such as protein localization, structure of the molecular machine complex, etc.

—————→ Strong link with the biostatistician community to build (for instance) specialized GSMM from omics data

Conclusion and perspectives

the RBA framework

- ❑ Extension of the RBA theoretical framework (under progress and/or validation)
 - ✓ To dynamical conditions (dRBA)
 - ✓ To stochastic fluctuations in gene expression
 - ✓ To include thermodynamics and kinetics constraints to predict the metabolite abundances
 - ✓ To predicts the emergence of regulatory networks
 - ✓ To the chemostat

- ❑ RBA for bioengineering to aid strain design

- ❑ Automatic generation of RBA models (RBApy)
 - ✓ Extension to eukaryotic cells
 - ✓ Integration of facilities for model manipulation, adaptation and visualization
 - ✓ Integration of additional methods of simulation (such as dRBA)

- ❑ Resource allocation for other prokaryotes and multi-cellular organisms (under progress and/or validation)
 - ✓ Microorganisms: *Escherichia coli*, *Streptomyces coelicolor*, *Ralstonia solanacearum*, *Synechocystis sp PCC6803*, yeast
 - ✓ Plant: *Arabidopsis thaliana*, *Zea mays*

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