Biological prior for network inference with Gaussian graphical models.

Application to Estrogen Receptor Status in Breast Cancer .

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Gene regulatory network

Which measure to use ?

- \blacktriangleright Correlation
	- Tends to group genes with close expression profiles

• Do not provide any clue on how the chain of information goes from gene to gene

\blacktriangleright Partial Correlation

• Quantify the correlation between two genes after excluding the effects of other genes

High dimensional setting

- \blacktriangleright "large p, small n" \rightarrow number of random variables (p) is much larger than the number of individuals (n)
- \triangleright p(p 1)/2 possible interactions
- Handling the scarcity of data
	- \blacktriangleright Sparsity:

Among all possible interactions only a few actually take place.

 \triangleright Coefficient matrix with mostly zero-valued entries

Regularized Gaussian graphical model

- \triangleright GGM: a well-studied framework to spot those direct relationships
- Dependency pattern described by the covariance matrix (independency between variables \Leftrightarrow absence of edge)
- \triangleright Sparse estimation via L1-regularization

A challenging issue

A vaste space of possible network structures

Biological prior knowledge could be used to limit the set of candidate networks

Method

a) Biological prior definition: differential and pathway analysis

b) Network inference: regularized GGM, multitask strategy

2 Application

- a) Context: ER status in Breast Cancer
- b) Results and interpretation

3 Conclusion

Differential analysis

 $\chi^{(\scriptscriptstyle C)}_{\!\! ig}$: expression level of the ith sample for gene g under condition c

$$
\mathbb{E}(X_{ig}^{(c)}) = \mu_g^{(c)} \quad \text{and} \quad \mathbb{V}(X_{ig}^{(c)}) = \sigma_g^2,
$$

Null hypothesis to test:

$$
\begin{cases}\nH_0: \quad \mu_g^{(1)} = \mu_g^{(2)}, \\
H_1: \quad \mu_g^{(1)} \neq \mu_g^{(2)}.\n\end{cases}
$$

Limma t-statistic (Smyth 2004)

$$
f_{\mathcal{G}}^{\text{limma}} = \frac{\bar{X}_{\mathcal{G}}^{(1)} - \bar{X}_{\mathcal{G}}^{(2)}}{S_{\mathcal{G}}^{\text{limma}} \sqrt{\frac{1}{n_1} + \frac{1}{n_2}}},
$$

- $\bullet \,$ $S_{\mathcal{G}}^{\text{\tiny{limma}}}$: Bayesian estimator of the variance
- Stabilize the estimation of gene variances $\frac{9}{735}$

Method - Summary

Microarray data

How to interpret gene signatures in biologically meaningful terms ?

 \rightarrow by determining whether the signature is enriched in pathway* key actors.

Figure: Group testing for pathway analysis

* Pathway: set of gene interacting in order to achieve a specific cellular function $11 / 35$

Under the null hypothesis of no over-representation

$$
\mathbb{P}(Y \geq y) = 1 - \mathbb{P}(Y \leq y)
$$

=
$$
1 - \sum_{i=0}^{y} \frac{\binom{s}{i} \binom{p-s}{t-i}}{\binom{p}{t}}.
$$

 $P(Y > y)$ probability of **observing at least y genes** of a **pathway of size t** in the signature

In practice...

Table: Results of pathway analysis

• Pathways do not clearly represent distinct entities ! \rightsquigarrow we need to summarize the set of pathways found significant

Method - Summary

Method Network Inference

R package SIMoNe : general settings

- I Enables inference of undirected networks:
	- \triangleright In a Gaussian graphical models (GGM) framework
	- \triangleright Multitask inference strategy: joint estimation of the graphs by coupling the estimation problems
- Based on partial correlation coefficients

Chiquet et al. 2010, Inferring Multiple Graphical Models. Statistics and Computing

Graphical model

Def.: Probabilistic model for which a graph denotes the conditional independence structure between random variables.

Gaussian model for an i.i.d. sample

- Let $P = \{1, ..., p\}$ be a set of nodes (i.e. genes)
- \blacktriangleright $X = (X_1, ... X_p)^T$ is the signal over this set (i.e. the gene expression levels), such as: $X \sim \mathcal{N}(\mathbf{0}_D, \Sigma)$
- \blacktriangleright Let Θ be the parameter to be inferred (i.e. the edges)

$$
\triangleright \Theta = (\theta_{ij})_{i,j \in \mathcal{P}} \triangleq \mathbf{\Sigma}^{-1} \text{ is the concentration matrix.} \triangleright \text{cor}_{ij|\mathcal{P}\setminus \{i,j\}} = -\theta_{ij}/\sqrt{\theta_{ij}\theta_{jj}} \text{ for } i \neq j
$$

Interpretation

If 2 nodes i and j are partially uncorrelated, no edge is inferred:

 $\lambda_i \perp\!\!\!\!\perp \lambda_j |X({\cal P} \backslash \{i,j\}) \Leftrightarrow \theta_{ij} = 0$

After a simple rescaling Θ can be interpreted as the adjacency matrix

conditional dependency or non null partial correlation between

Method - Network Inference

Let $S = n^{-1}X^{T}X$ be the empirical variance-covariance matrix.

- \blacktriangleright **S**⁻¹ is not defined for $n < p$.
- If $n < p$, neither Θ nor its support can be estimated
- \blacktriangleright The need for regularization is huge

Estimation: a penalized likelihood approach

$$
\hat{\Theta}_{\lambda} = \underset{\Theta}{\text{arg}\max} \mathcal{L}(\Theta; \text{data}) - \lambda \ \text{pen}_{\ell_1}(\Theta),
$$

- \triangleright $\mathcal L$ is the model log-likelihood,
- $\blacktriangleright \ \text{pen}_{\ell_1} = ||\Theta||_{\ell_1}$ is a penalty function tuned by $\lambda > 0$.

It performs:

- 1 regularization (needed when $n \ll p$),
- 2 selection (sparsity induced by the ℓ_1 -norm)

Take into account the core-pathways information as an *a-priori* knowledge:

 \rightsquigarrow Edges between two genes of the same core-pathway are less penalized

Statistical approach

Use adaptive penalty parameters for different coefficients

 \blacktriangleright Let Z be the set of indicator variable for nodes

$$
\hat{\Theta}_{\lambda} = \arg \max_{\Theta} \mathcal{L}(\Theta; \text{data}) - \lambda \|\mathbf{P}_{\mathbf{Z}} \star \Theta\|_{\ell_1},
$$

where P_z is a matrix of weights depending on the core-pathway membership Z.

Method - Network Inference

Multitask inference

 \rightarrow How to deal with various conditions?

- \triangleright Assumption: strong relationship between both networks
- \triangleright Approach: joint estimation of the graphs by coupling the estimation problems

Consider C conditions where the same p genes are measured

Graphical coop-LASSO

$$
\begin{aligned}\n\max_{\Theta^{(c)}} \sum_{c=1}^{C} & \mathcal{L}\left(\Theta^{(c)};\text{data}\right) \\
&\quad - \lambda \sum_{\substack{i,j \in \mathcal{P} \\ i \neq j}} \left\{ \left(\sum_{c=1}^{C} \left[\theta_{ij}^{(c)} \right]_{+}^{2} \right)^{1/2} + \left(\sum_{c=1}^{C} \left[\theta_{ij}^{(c)} \right]_{-}^{2} \right)^{1/2} \right\},\n\end{aligned}
$$

where $[u]_{+} = max(0, u)$ and $[u]_{-} = min(0, u)$.

- \blacktriangleright Group-lasso like penalty
- Disconnect the activation of up and down regulation

Method - Network Inference

 $\varphi \mathcal{Q} = \{1, \ldots, \mathsf{Q}\}\$ of given overlapping core-pathways \triangleright Z_{iq} = 1 if $i \in q$ and 0 otherwise

Maximisation Problem

$$
\max_{\theta^{(c)}} \sum_{c=1}^{C} \mathcal{L}\left(\Theta^{(c)};\text{data}\right) - \lambda \sum_{\substack{i,j \in \mathcal{P} \\ i \neq j}} \rho_{\mathbf{Z},\mathbf{Z}_j} \left\{ \left(\sum_{c=1}^{C} \left[\theta_{ij}^{(c)} \right]_{+}^{2} \right)^{1/2} + \left(\sum_{c=1}^{C} \left[\theta_{ij}^{(c)} \right]_{-}^{2} \right)^{1/2} \right\}, \quad (1)
$$

where $[u]_{+}$ = max(0, u) and $[u]_{-}$ = min(0, u) and the coefficients of the penalty are defined as:

$$
\rho_{\mathbf{Z},\mathbf{Z}_j} = \begin{cases}\n\sum_{q,\ell \in \mathcal{Q}} Z_{iq} Z_{j\ell} \frac{1}{\lambda_{in}}, & \text{if } i \neq j, \text{ and } q = \ell, \\
\sum_{q,\ell \in \mathcal{Q}} Z_{iq} Z_{j\ell} \frac{1}{\lambda_{out}}, & \text{if } i \neq j, \text{ and } q \neq \ell, \\
1, & \text{otherwise.} \n\end{cases}
$$
\n(2)

Method - Summary

Application ER status in breast cancer

Breast cancer in a few words

 \triangleright An heterogeneous disease (5 subtypes)

Presence $(ER+)$ /absence $(ER-)$ of estrogen receptors: an essential parameter of tumor characterization.

 \sim Understanding the molecular mechanism of ER status: a key issue for treatment and prognosis

Inference of regulation networks under ER+ and ERconditions

• Comparison of regulation patterns

Protein trafficking

Small molecules biochemistry

Figure: Core pathways

Figure: Sub-network inferred from the ER status signature

Anti-apoptotic mechanisms

Common regulations

Estrogen receptor (ESR1) - BCL2 (Peterson at al. 2007)

ESR1 - EGFR/IGF1R (Salvatori et al. 2000, Oesterreich et al. 2001)

Specific regulations

EGF receptor family: ERBB3 - ERBB4 (Lee et al. 2001) CDK6 - IGF1R

Figure: Anti-apoptotic mechanisms

Summary

- \blacktriangleright Very challenging issue
- \blacktriangleright Introducing biological priors reduce the space of possible networks
- **Promising application on Breast cancer dataset**
- \blacktriangleright Importance of missing covariates

 \rightsquigarrow Persepectives: need for integration of heterogeneous omics data.

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